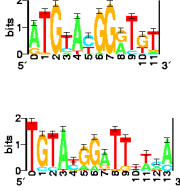
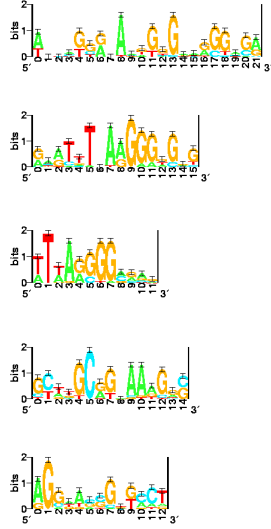
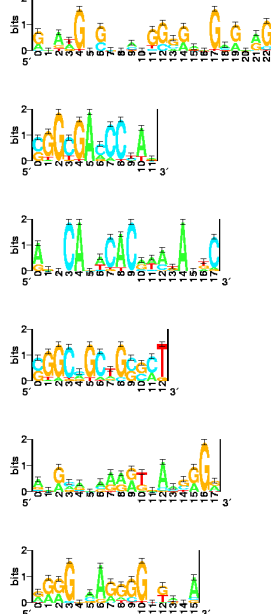


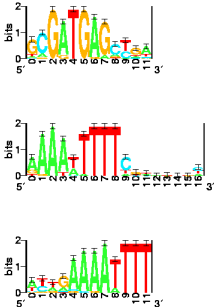
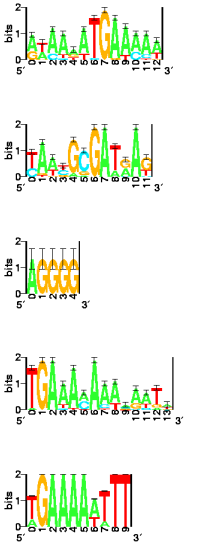
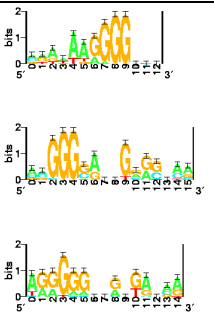
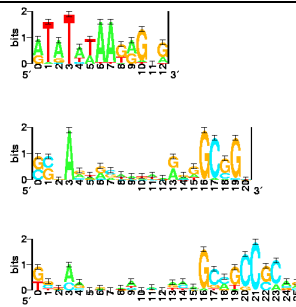
## Supplemental Information

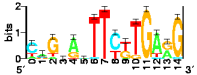
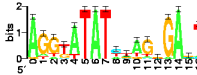
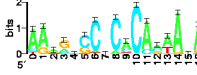





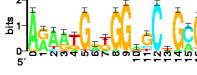

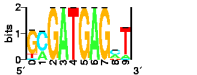




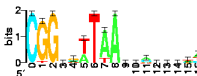
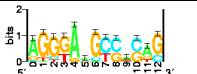




**Includes: Supplemental Table I, Supplemental References, Supplemental Figure 1, Supplemental Figure 2A-D, Supplemental Figure 3.**

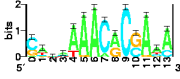

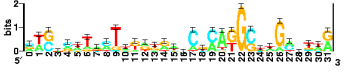

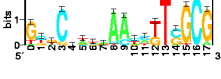



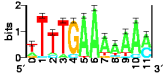


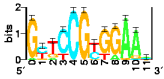




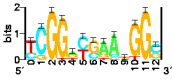



**Prevalence of sequence features chosen by the Bayesian network.** The types of constraints learned by the network indicate the prevalence of various modes of combinatorial regulation in *S. cerevisiae*. It is important to note that the degree of combinatorial regulation uncovered here represents a lower limit, and a more diverse sampling of physiological conditions is likely to yield a higher average number of regulators per gene, and more complex combinatorial rules. Of 6,906 parents selected on 5 cross-validations and 10 re-samplings on 49 clusters, 6,498 constraints (94%) were for the presence or absence of a motif. Constraining the distance to ATG was chosen 5.1% of the time, for subset of motifs, foremost among which are PAC, RRPE, RPN4, STRE, and RAP1, indicating potential mechanistic constraints for their function. Orientation was constrained 0.6% of the time, and the vast majority of these were for RAP1. So our analysis indicates that transcriptional regulation by RAP1 depends on its orientation, but that most other factors, including PAC and RRPE, work independently of orientation. All other constraints were chosen at even lower frequencies. While the learned sequence features are predictive, some functional constraints may not be detected if they are regularly satisfied in a set of genes selected by a previous constraint.

**Supplemental Table I.** High confidence motifs learned from the Bayesian Network for each cluster, found in at least 25% of the re-samplings.

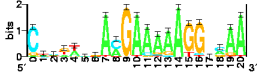

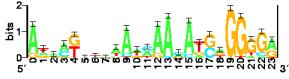
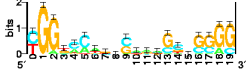

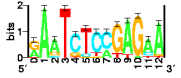



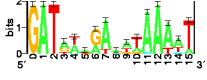

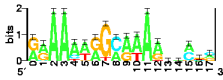


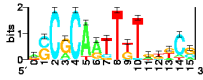
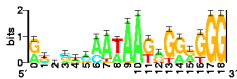

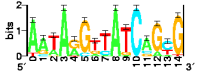



Cluster	Known motifs	New motifs
1	 <p>RAP1</p> <p>RAP1-var</p>	
2		 <p>STRE-like</p> <p>STRE-like</p>
3		 <p>STRE-like</p>

4	 <p>PAC</p> <p>RRPE</p> <p>RRPE</p>	
5	 <p>HSF1-var</p>	
6	 <p>STE12-like</p> <p>PAC-var</p> <p>STRE</p> <p>STE12</p> <p>RRPE</p>	
7	 <p>STRE</p> <p>STRE-like</p> <p>STRE</p>	 <p>TATA-var</p>

8	 <p>HSF1-like</p>	   
9	 <p>MCM1-like</p>  <p>REB1-like</p>	  
10	 <p>PAC</p>  <p>PAC-var</p>  <p>PAC</p>	
11	 <p>RPN4</p>  <p>OAF1</p>	    

12	 <p>bits 5' 3'</p> <p>SWI4</p>	 <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>
13	 <p>bits 5' 3'</p> <p>STE12-like</p>	 <p>bits 5' 3'</p>  <p>bits 5' 3'</p>
14	 <p>bits 5' 3'</p> <p>PDR3</p>	 <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>
15	 <p>bits 5' 3'</p> <p>MIG1</p>	 <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>  <p>bits 5' 3'</p>

16		
17		<p>ABF1-like</p> <p>RRPE-var</p> <p>RRPE</p> <p>REB1-like</p>
18		
19		<p>STRE</p>

		
20	 <p>HAP4</p>	  
21	 <p>HSF1-like</p>  <p>RPN4-like</p>	 
22		 
23		  
24	 <p>MET4</p>  <p>STRE</p>	    


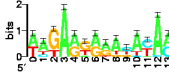




		RRPE-var	
30	 	MBP1 MBP1-var	
31	 	INO4-like SWI4-like	
32		PAC-like	   
33		STRE-var	  
34			
35			 

36	<p>RAP1-like</p>	
37		
38	<p>YAP1</p>	
39		

40			
41			
42			
43			
44			
45		MCM1	
46		HSF1	
47			
48			

49		GCR1	
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**Supplemental References for motifs learned by the network which have been previously documented in the literature:**

RAP1: Planta, R.J., et al, *Biochem. Cell Biol.* (1995); Warner, J. *Trends Biochem. Sci.* (1999); Lieb, J.D., et al, *Nat. Genetics* (2001).

RRPE (M3a): Tavazoie, S., et al, *Nat. Genetics* (1999).

PAC (M3b): Tavazoie, S., et al, *Nat. Genetics* (1999).

HSF1: Morimoto, R.I., *Science* (1993).

STRE (MSN2/4): Martinez-Pastor, M.T., et al, *EMBO J* (1996).

UME6: Strich, R., et al, *Genes Dev.* (1994).

MET4: Thomas, D. & Surdin-Kerjan, Y. *Microbiol. Mol. Biol. Rev.* (1997).

MCM1: McIney, C. J., et al, *Genes Dev.* (1997)

REB1: Morrow, B. E., et al, *JBC* (1989); Liaw, P.C.Y., et al., *Yeast* (1994).

RPN4: Mannhaupt, G., et al, *FEBS Lett.* (1999); Jelinsky, S.A., et al, *MCB* (2000).

OAF1: Rottensteiner, H., et al, *Eur. J. Biochem.* (2003).

SWI4: Koch, C, and Naysmith, K. *Curr. Opin. Cell Biol.* (1994).

PDR3: Delahodde, A., et al, *MCB* (1995).

MIG1: Lundin, M., et al, *MCB* (1994).

ABF1: Gray, W.M. and Fassler, J.S, *Gene Expr.* (1993); Planta, R.J., et al, *Biochem. Cell Biol.* (1995).

HAP4: Forsburg, S. L., and Guarente, L., *Genes Dev.* (1989).

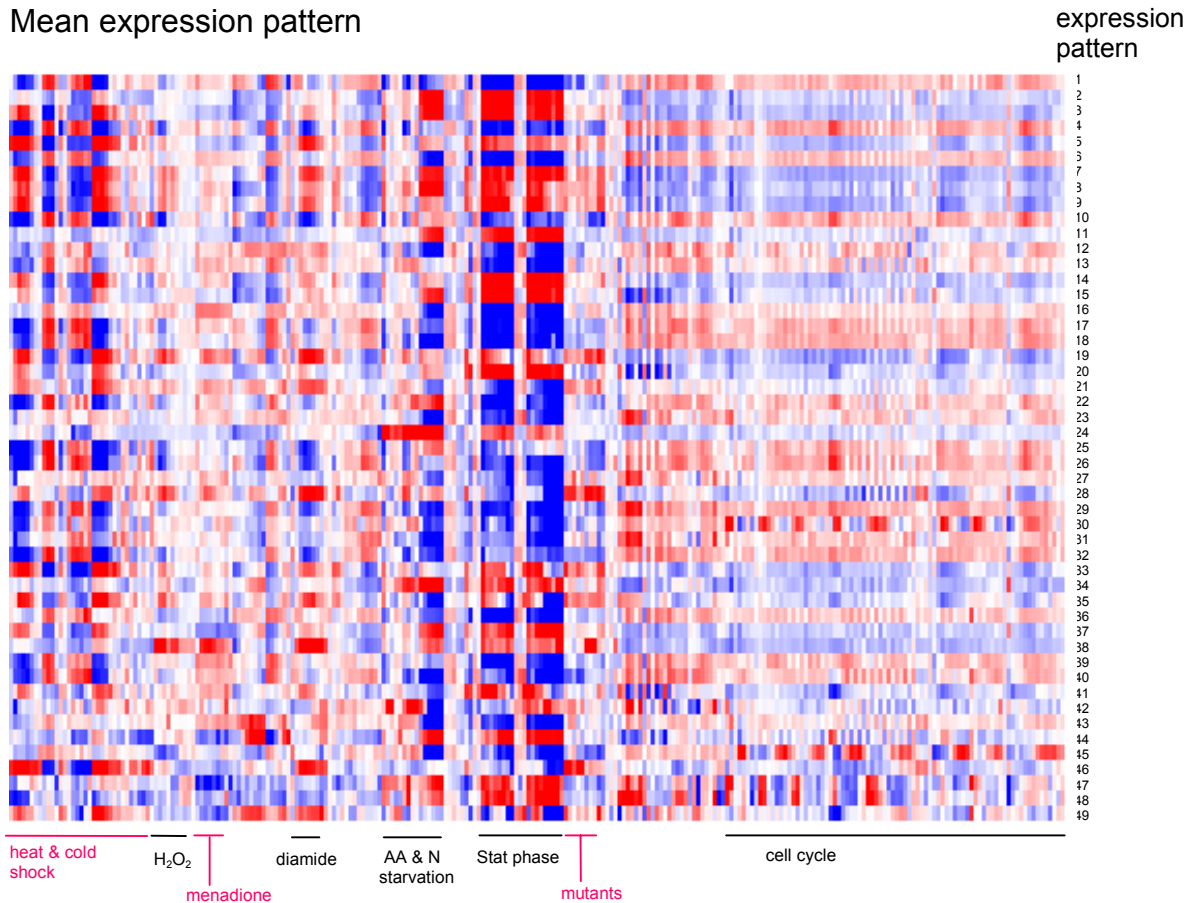
CBF1: Thomas, D. & Surdin-Kerjan, Y. *Microbiol. Mol. Biol. Rev.* (1997).

MBP1: Macintosh, E.M., et al, *Curr. Gen.* (1993); Koch, C, and Naysmith, K. *Curr. Opin. Cell Biol.* (1994).

STE12: Baur, M., et al, *MCB* (1997).

YAP1: Fernandes, L., et al, MCB (1997).

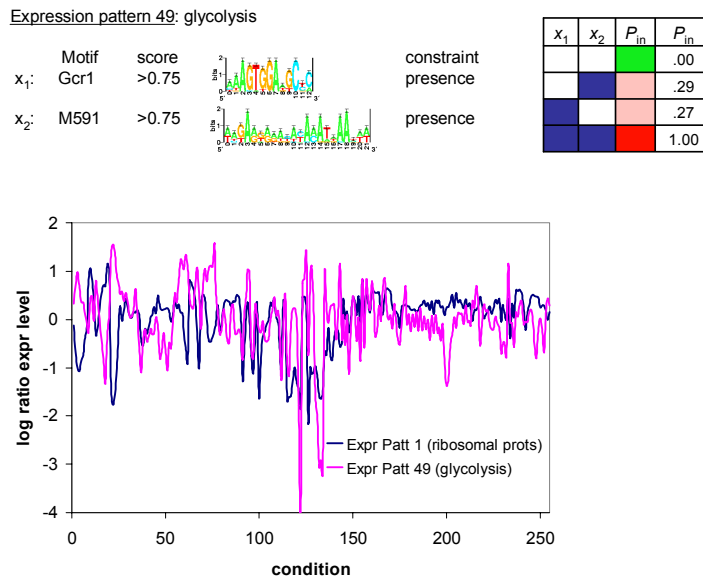
GCR1: Deminoff, S.J., et al, Genetics (2001).



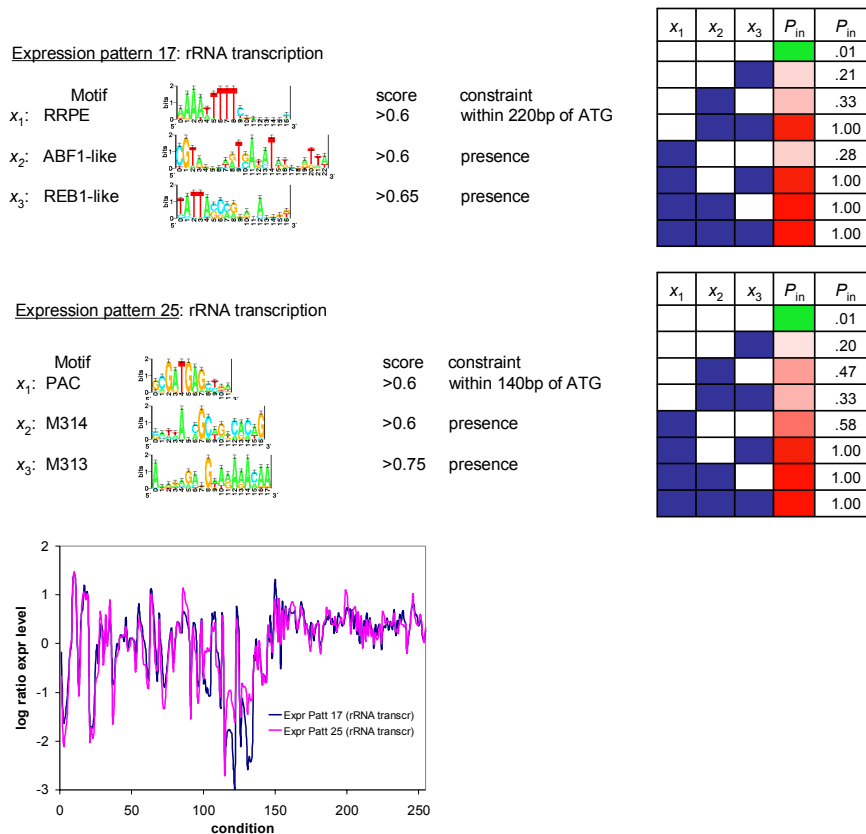
**Supplemental Figure 1.** Mean expression of all 49 expression patterns. Particularly informative conditions are highlighted. While the two main responses are stress induced and stress repressed, there are subtle differences between expression patterns which can distinguish functionally distinct but similarly expressed sets of genes. For example, expression pattern (1) is almost exclusively ribosomal protein genes, while the similar expression pattern (4) is highly enriched for genes involved in ribosomal RNA transcription and processing. Expression patterns (30), (45), (47) and

(48) are cell cycle regulated genes, but they remain significantly co-regulated across the environmental stress conditions.

**Supplemental Figure 2.** Examples of expression patterns explained by the network. Here we select subsets of the constraints found by the full network, to highlight prevalent modes of combinatorial regulation and biologically interesting examples.

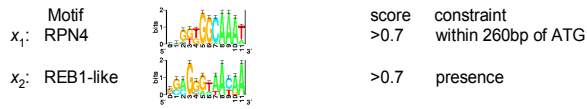


**Supplemental Fig 2A.** Glycolysis, expression pattern 49. Top: The network for expression pattern 49 learns Gcr1 and a new motif M591. Genes with only one are in the expression pattern 29% and 27% of the time, but genes with both motifs are in the expression pattern 100% of the time. Bottom: Expression pattern of this set compared to the ribosomal protein expression pattern (1). These GCR1 regulated genes are anti-correlated with the ribosomal proteins over many conditions.



**Supplemental Fig 2B.** rRNA transcription, expression patterns 17 and 25. These expression patterns are similar to expression pattern 4, the largest rRNA transcription expression pattern (regulated by PAC and RRPE, Fig 3A in main text). But here RRPE and PAC are learned in combination with other motifs. Top: In expression pattern 17, the network learns RRPE with new motifs similar to the known binding sites for ABF1 and REB1. Expression pattern 25 learns PAC in combination with the new motifs M314 or M313. Bottom: Expression pattern 17 and 25 are similar, with subtle differences in the degree of repression under amino acid starvation and stationary phase. Expression pattern 4 falls between these.

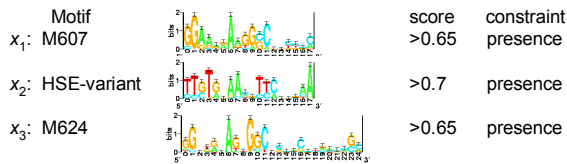
Expression pattern 28: proteolytic degradation



$x_1$	$x_2$	$P_{in}$	$P_{in}$
		.00	
	.19	.19	
.46		.46	
1.00	1.00	1.00	

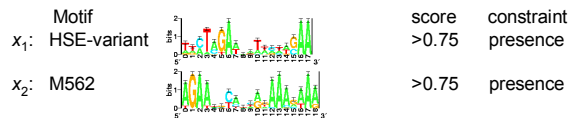
**Supplemental Fig. 2C.** Proteolytic degradation, expression pattern 28. When RPN4 is constrained to be within 260bp of ATG, genes are in the expression pattern 46% of the time. With the additional presence of a REB1-like motif, the fraction is 100%.

Expression pattern 5: C-compound metabolism

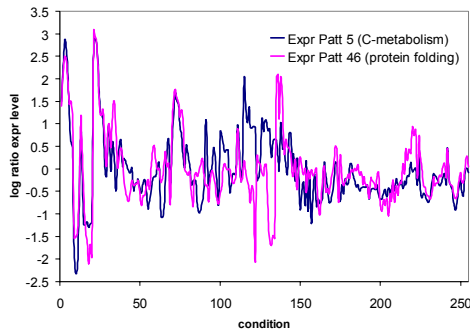


$x_1$	$x_2$	$x_3$	$P_{in}$	$P_{in}$
			.01	
		.69	.69	
	.42		.42	
			1.00	
.72			.72	
1.00			1.00	

Expression pattern 46: protein folding

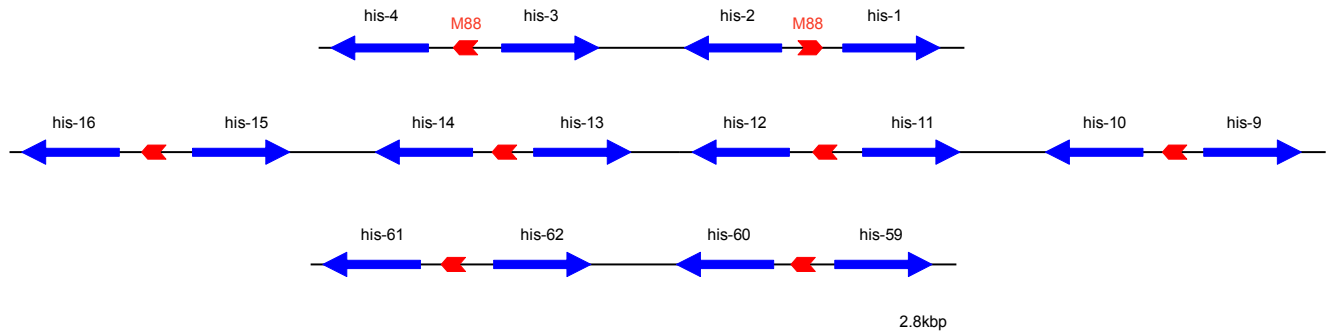


$x_1$	$x_2$	$P_{in}$	$P_{in}$
		.00	
	.15	.15	
.25		.25	
.93		.93	



**Supplemental Fig. 2D.** C-compound metabolism and protein folding, expression patterns 5 and 46. These expression patterns are both induced under heat shock but are anti-correlated during stationary phase, where the protein folding genes are repressed but C-compound metabolism genes are induced. Networks for both expression patterns learn variants of the heat shock element (HSE), but with different regulatory partners.





**Supplemental Fig 3.** Examples of *C. elegans* histone genes selected by the network for expression pattern 15. All histones are selected by motif M88.